



# Construction of mutant library V.2

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Version 2 ▼

1 Works for me

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## ABSTRACT

This protocol is used to construct mutant library of target gene with high efficiency and low false positives/negatives rate after subsequent functional screening.

## DOI

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## PROTOCOL CITATION

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## MATERIALS TEXT

PCR tube, thermocycler, ddH<sub>2</sub>O, Nanodrop

Error-prone PCR:

Random Mutagenesis Kit by Solarbio

Plasmid template

MEGAWHOP PCR:

2×High Fidelity Master Mix

DpnI digestion:

DpnI (NEB) (20,000units/ml)

10xCutsmart

Product purification

E.Z.N.A.® Cycle Pure Kit

## SAFETY WARNINGS

Please wear gloves for the experiment, don't try to touch the lid after PCR program initiation.

## BEFORE STARTING

Make sure that the template of MEGAWHOP PCR is fresh to improve the construction efficiency.

## Error-prone PCR

- 1 Add the following reagent to a PCR tube (50µl) (Random Mutagenesis Kit by Solarbio).

<b>A</b>	<b>B</b>
Template(10µl)	Depends on the concentration
Forward Primer (10 µM)	1 µl
Reverse Primer (10 µM)	1 µl
Mut Enhancer	3µl
2 x Mut Random System	25µl
ddH <sub>2</sub> O	Add to 50µl

- 2 Program the thermocycler as follows:

A	B
Temperature	Time
95°C	2min
94°C	30 s
Tm-3~5°C	1min
72°C	1kbp/min
72°C	7 min
16°C	∞

- 3 Use the palm centrifuge to mix the solution in PCR tube.
- 4 Put the PCR tube into the thermocycler and Run the program.
- 5 Using agarose gel electrophoresis to confirm if correct construct was present.

#### PCR product purification

- 6 PCR product purified by E.Z.N.A.® Cycle Pure Kit.
- 7 Test the concentration and purity of DNA using NanoDrop.

#### MEGAWHOP PCR

- 8 Add the following reagent to a PCR tube (50µl).

A	B
Plasmid template (50µl)	Depends on the concentration
Purified Production of error-prone PCR (mega primer) (500µl)	Depends on the concentration
2×High Fidelity Master Mix (Enzyme)	25 µl
ddH2O	Add to 50µl

- 9 Program the thermocycler as follows:

A	B
Temperature	Time
95°C	5min
95°C	30s
Depends on the Tm	30s
72°C	2kb/min
72°C	7 min
16°C	∞

- 10 Use the palm centrifuge to mix the solution in PCR tube.
- 11 Put the PCR tube into the thermocycler and Run the program.
- 12 Using agarose gel electrophoresis to confirm if correct construct was present.

#### PCR product purification

- 13 PCR product purified by E.Z.N.A.® Cycle Pure Kit.
- 14 Test the concentration and purity of DNA using NanoDrop.

#### Dpnl digestion

- 15 Add the following reagents to a PCR tube (e.g. 20µl).

A	B
Dpnl (NEB) (20,000units/ml)	Depends on the quality of DNA (20units Dpnl digests 1µg DNA)
10xCutsmart	2µl
Purified Production of MEGAWHOP PCR	Moderate (e.g.400 ng)
ddH2O	Add to 20µl

- 16 Use the palm centrifuge to mix the solution in PCR tube.

17 Incubate at 37°C for 2 hours and heat inactivation 80°C for 20 min.

#### Digestion product purification

18 Digestion product purified by E.Z.N.A.® Cycle Pure Kit.

19 Test the concentration and purity of DNA using NanoDrop.

#### Nick ligation (T4 ligase)

20 Add the following reagents to a PCR tube (e.g. 20µl)

A	B
T4 DNA ligase (Thermo Fisher) (Weiss U)	1U
Purified Production of DpnI digestion	50ng
10X T4 DNA Ligase Buffer (Thermo Fisher)	2µl
ddH <sub>2</sub> O	Add to 20µl

21 Use the palm centrifuge to mix the solution in PCR tube.

22 Incubate the reaction at 16°C overnight.

#### Transformation

23 Transform the nick ligation product into competent cells.